

DOI: 10.21767/2574-285X.100008

# The Potential of Infrared Spectroscopy and Multivariate Analysis of Peripheral Blood Components as a Validated Clinical Test for Early Diagnosis of Alzheimer's Disease

**Salman A<sup>1</sup> and Mordechai S<sup>2</sup>**<sup>1</sup>Department of Physics, SCE - Shamoon College of Engineering, Beer-Sheva 84100, Israel<sup>2</sup>Department of Physics, Ben-Gurion University of the Negev, Beer-Sheva 84105, Israel**Corresponding authors:** Dr. Ahmad Salman, Department of Physics, SCE - Shamoon College of Engineering, Beer-Sheva 84100, Israel, Tel: 97286475794; Fax: 97286475758; E-mail: ahmad@sce.ac.il

Prof. Shaul Mordechai, Department of Physics, Ben-Gurion University of the Negev, Beer-Sheva 84105, Israel, Tel: 97286461749; Fax: 97286472903; E-mail: shaulm@bgu.ac.il

**Received:** December 26, 2016; **Accepted:** December 30, 2016; **Published:** January 05, 2017**Citation:** Salman A, Mordechai S. The Potential of Infrared Spectroscopy and Multivariate Analysis of Peripheral Blood Components as a Validated Clinical Test for Early Diagnosis of Alzheimer's Disease. Cell Mol Med 2017, 2:1.

## Opinion

Alzheimer's disease (AD) is usually considered as an aging disease as the Greek and Roman physician used to consider, and as the main cause of dementia. AD patients start to do mistakes in language, to lost direction, changes in mood, self-careless until losing their emotional and body functions until death [1]. The symptoms of the disease are getting worse by time. AD was first diagnosed in 1901 by psychiatrist Alois Alzheimer (thus this dementia acquired his name) and till now its causes are not yet known and there is no validated clinical test for diagnosis [2] especially at early stages. According to the statistics in 2015, there is about 48 million persons worldwide diseased with AD [3]. In 2010, 486,000 deaths resulted from dementia. As life expectation increases in developed countries and AD is mainly an aging disease it becomes financially very costly disease [4]. While the death due to other diseases has been decreased, the deaths related to AD are increasing; for example the deaths due to AD in the years 2000 and 2008 have increases by 66%. The economic burden due to AD is very high making it a major medical challenge; the cost of the disease in the USA alone in 2015 was \$226 billion and it is in the upward trend [5].

Although there is no medication to treat or reverse the disease, it is important to diagnose AD accurately as early as possible, since there are medications to slow the symptoms and thus to improve the quality of life by preserving daily functioning for more time [3]. These medications are most effective when used at the early stages of AD.

The initiation of the disease is associated with the formation of senile plaques, mitochondrial dysfunction, and hyper phosphorylated tau protein in neurofibrillary tangles in the brain [6].

Today, AD is diagnosed subjectively by a combination of cognitive neurologists, geriatricians, and psychiatrists based on medical files and mental tests such as mini-mental state examination (MMSE). Medical imaging such as positron

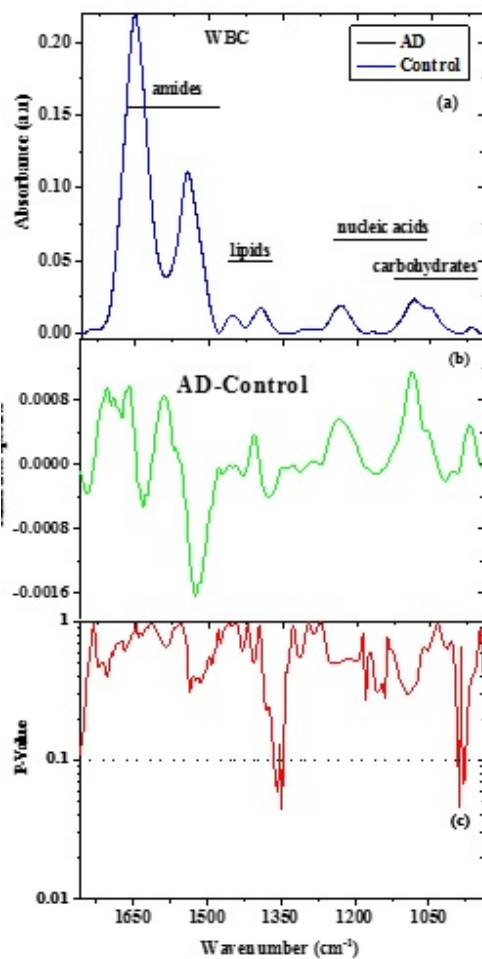
emission tomography (PET) and magnetic resonance imaging (MRI), computerized tomography (CT) [7] and Cerebrospinal fluid (CSF) [8] are combined to the cognitive tests to enable reliable diagnosis. Actually, it is possible to diagnose AD accurately only just after death [7,9] moreover, 45% of AD patients have not been told about their disease.

Many new techniques were developed for AD diagnosis like, several CSF biomarkers tau, amyloid beta [10], single photon-emission computed tomography (SPECT) [11], spectral method for CSF biomarker analysis (ELISA) [12], mass spectrometry, Raman, polymerase-chain-reaction (PCR)). Different studies used brain tissues for AD diagnosis such as Raman [13] and FTIR microscopy [14] and near infrared fluorescence probes [15].

There are major drawbacks of these new techniques, for example, CSF biomarkers are not well standardized (sample handling, concentration) [16]; the pathognomonic and diagnostic value of the genetic and biochemical markers are not proofed yet (expensive equipments and it is hazardous to extract the CSF [17]). All these drawbacks and the situation of AD early diagnosis empathize the need of objective and reliable technique for clinic diagnosis and follow up of patients which can be used routinely [17]. These features of the diagnostic technique make the blood test as the most promising biomarker [18].

Infrared spectroscopy is known as a cheap, rapid and easy to use technique for objective analysis. It is widely used in the medical fields [19-24] for more than three decades for the detection of cancerous disease based on biomedical changes occurring during the developing of the disease related to the alteration in the chemical composition [25].

Mid-infrared absorption spectra are obtained using infrared spectroscopy by measuring the absorption bands of the functional groups of the molecules that compose the examined sample. These spectra can be used for qualitative and quantitative analysis and is considered as a fingerprint of the biological samples.



**Figure 1** Average infrared absorption spectra (a), difference spectra (b) and p-value spectra (c) of WBC for AD patients and controls in the region 942-1760  $\text{cm}^{-1}$ .

Recently infrared spectrometers have benefitted from high spectral and spatial resolution developments making the infrared spectroscopic technique as a reliable in both applied and basic research. In addition, there was a great development of new bioinformatics methods for the analysis of bio-fluid samples from patients with different diseases [26].

Combining bioinformatics methods in tandem with infrared microscopy generate a powerful diagnostic tool for health care using the analysis of bio fluids like serum [27], urine, plasma and white blood cells based on their infrared absorption spectra [28]. There are many advantaged of bio fluids analysis since they are minimal invasive and cheap which makes it ideal for routine and low risk method for clinics [28].

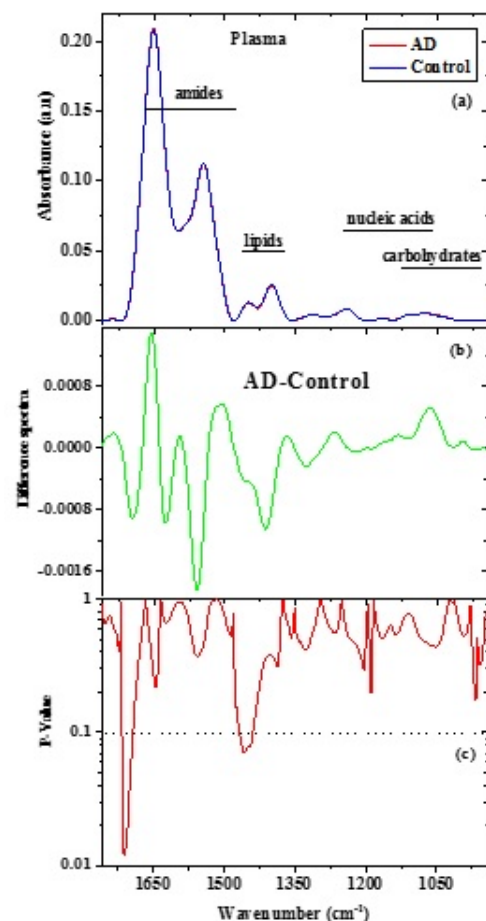
Many studies have been carried out and showed the potential of infrared microscopy of blood components in the medical field [29,30]. For example, detection of different kinds of cancers, detection of AD based on plasma samples and the analyses of the secondary structure due to amyloids beta [31].

Different biochemical changes are associated with AD and just part of them are known like amyloid beta peptides,

changes in lymphocytes in AD patients [32], but most of these changes are still not known [32].

Several studies have shown the potential of FTIR microscopy in tandem with multivariate analysis for the early detection of AD [33] and recently our group have shown the potential of this spectroscopy technique for early diagnosis of AD using WBC [34]. There is clear evidence of the ability of multivariate analysis for classification manner even if the changes among the groups are minute [35-39].

**Figures 1a and 2a** show the examples of mid infrared spectra for two of the blood components WBC and plasma showing the difference between groups. As expected, the spectra are generated as results of the vibrations of the functional groups of the biomolecule that compose the biological samples, such as proteins, lipids carbohydrates and nucleic acid.



**Figure 2** Average infrared absorption spectra (a), difference spectra (b) and p-value spectra (c) of plasma for AD patients and controls in the region 942-1760  $\text{cm}^{-1}$ .

The differences between the healthy control and AD patients in both WBC and plasma samples are subtle but they are still sufficiently repeatable to yield the required statistics. The difference spectra calculated as AD minus control for WBC and plasma are plotted in **Figures 1b and 2b** respectively. **Figures 1c and 2c** show the p-values calculated using t-test to

differentiate between the two groups based on WBC and plasma samples respectively. The calculations were performed for each wavenumber separately. As can be seen from the figures there are several sensitive wavenumbers which enable to differentiate significantly (>90%) between AD patients and controls. In the plasma the bands centered at 1713  $\text{cm}^{-1}$  attributed to amide I (C=O, thymine), and the wavenumbers in the range 1440-1469  $\text{cm}^{-1}$  (due to lipids and proteins) [40]. Both wavenumbers can be considered as biomarkers, which differentiate between AD patients and controls. In WBC, the most sensitive wavenumbers are found in the regions 980-995  $\text{cm}^{-1}$  (OCH<sub>3</sub> polysaccharides and C-O ribose [41] and C-C) and 1347-1368  $\text{cm}^{-1}$  (arising due to C-O stretching C-H and N-H deformations).

Using multivariate analysis PCA followed by LDA it was possible to differentiate between AD regardless of the disease stage and control with more than 90% success rate [34].

Our results indicate that the diagnosis of AD at early stages is plausible based on simple peripheral blood tests. Infrared microscopy combined with machine learning algorithms makes this method more reliable and objective as a diagnostic tool.

## References

1. A.A.s.D.S.N.I.o.A.R.D. (2011).
2. Strassnig M, Ganguli M (1907) About a peculiar disease of the cerebral cortex. By Alois Alzheimer, 1907 (Translated by L. Jarvik and H. Greenson), *Alzheimer disease and associated disorders* 1: 3-8.
3. D.F.s.N.W.H.O.M.A.ft.o.o.M.R. January (2016).
4. Bonin-Guillaume S, Zekry D, Giacobini E, Gold G, Michel JP (2005) The economical impact of dementia, *Presse medicale* (Paris, France : 1983) 34: 35-41.
5. Alzheimer's Association (2015) Alzheimer's disease facts and figures. Retrieved on August 11.
6. Ballard C, Gauthier S, Corbett A, Brayne C, Aarsland D (2011) Alzheimer's disease, *Lancet* 377: 1019-1031.
7. <http://www.mayoclinic.org/diseases-conditions/alzheimers-disease/diagnosis-treatment/diagnosis/dxc-20167109>, in.
8. Andreasson U, Vanmechelen E, Shaw LM, Zetterberg H, Vanderstichele H (2012) Analytical aspects of molecular Alzheimer's disease biomarkers. *Biomark Med* 6: 377-389.
9. <https://www.nia.nih.gov/alzheimers/topics/diagnosis>, in.
10. Blennow K, Hampel H, Weiner M, Zetterberg H (2010) Cerebrospinal fluid and plasma biomarkers in Alzheimer disease. *Nat Rev Neurol* 6: 131-144.
11. <http://www.nhs.uk/Conditions/Alzheimers-disease/Pages/Diagnosis.aspx>.
12. Mori H, Hosoda K, Matsubara E, Nakamoto T, Furiya Y, et al. (1995) Tau in cerebrospinal fluids: Establishment of the sandwich ELISA with antibody specific to the repeat sequence in tau. *Neuroscience Letters* 186: 181-183.
13. Michael R, Otto C, Lenferink A, Gelpi E, Montenegro GA (2014) Absence of amyloid-beta in lenses of Alzheimer patients: A confocal Raman microspectroscopic study. *Experimental Eye Research* 119: 44-53.
14. Benseny-Cases N, Klementieva O, Cotte M, Ferrer I, Cladera J (2014) Microspectroscopy ( $\mu$ FTIR) reveals Co-localization of lipid Oxidation and amyloid plaques in human Alzheimer disease brains. *Analytical Chemistry* 86: 12047-12054.
15. Tong H, Lou K, Wang W (2015) Near-infrared fluorescent probes for imaging of amyloid plaques in Alzheimers disease. *Acta Pharmaceutica Sinica B* 5: 25-33.
16. Mattsson N, Zegers V, Andreasson U, Bjerke M, Blankenstein MA (2012) Reference measurement procedures for Alzheimer's disease cerebrospinal fluid biomarkers: definitions and approaches with focus on amyloid  $\beta$ 42. *Biomarkers in Medicine* 6: 409-417.
17. Carmona P, Molina M, Toledano A (2016) Blood-based biomarkers of Alzheimer's disease: Diagnostic algorithms and new technologies. *Current Alzheimer Research* 13: 450-464.
18. O'Bryant SE, Xiao G, Barber R (2010) A serum protein-based algorithm for the detection of alzheimer disease. *Archives of Neurology* 67: 1077-1081.
19. Mantsch H, Chapman D (1996) *Infrared spectroscopy of biomolecules*. Wiley-Liss, New York.
20. Zwielly A, Mordechai S, Sinielnikov I, Salman A, Bogomolny E (2010) Advanced statistical techniques applied to comprehensive FTIR spectra on human colonic tissues. *Med Phys* 37: 1047-1055.
21. Argov S, Ramesh J, Salman A, Sinielnikov I, Goldstein J (2002) Diagnostic potential of Fourier-transform infrared microspectroscopy and advanced computational methods in colon cancer patients. *J Biomed Opt* 7: 248-254.
22. Bellisola G, Sorio C (2012) Infrared spectroscopy and microscopy in cancer research and diagnosis. *American Journal of Cancer Research* 2: 1-21.
23. Salman A, Erukhimovitch V, Talyshinsky M, Huleihil M (2002) FTIR spectroscopic method for detection of cells infected with herpes viruses. *Biopolymers* 67: 406-412.
24. Lee-Montiel FT, Reynolds KA, Riley MR (2011) Detection and quantification of poliovirus infection using FTIR spectroscopy and cell culture. *Journal of Biological Engineering* 5: 16.
25. Chiriboga L, Xie P, Yee H, Zarou D, Zakim D (1998) Infrared spectroscopy of human tissue. IV. Detection of dysplastic and neoplastic changes of human cervical tissue via infrared microscopy. *Cell Mol Biol (Noisy-le-grand)* 44: 219-229.
26. Caine S, Heraud P, Tobin MJ, McNaughton D, Bernard CCA (2012) The application of Fourier transform infrared microspectroscopy for the study of diseased central nervous system tissue *NeuroImage* 59: 3624-3640.
27. González-Domínguez R, García-Barrera T, Gómez-Ariza JL (2015) Metabolite profiling for the identification of altered metabolic pathways in Alzheimer's disease. *Journal of Pharmaceutical and Biomedical Analysis* 107: 75-81.
28. Baker MJ, Hussain SR, Lovergne L, Untereiner V, Hughes C (2016) Developing and understanding biofluid vibrational spectroscopy: A critical review. *Chemical Society Reviews* 45: 1803-1818.
29. Barlev E, Zelig U, Bar O, Segev C, Mordechai S, et al. (2016) A novel method for screening colorectal cancer by infrared spectroscopy of peripheral blood mononuclear cells and plasma. *J Gastroenterol* 51: 214-221.

30. Zelig U, Barlev E, Bar O, Gross I, Flomen F (2015) Early detection of breast cancer using total biochemical analysis of peripheral blood components: A preliminary study. *BMC Cancer* 15: 408.
31. Nabers A, Ollesch J, Schartner J, Kötting C, Genius J (2016) Amyloid- $\beta$ -secondary structure Distribution in cerebro-spinal fluid and blood measured by an immuno-infrared-sensor: A biomarker candidate for Alzheimer's disease. *Analytical Chemistry* 88: 2755-2762.
32. Leandro GS, Lobo RR, Oliveira DV, Moriguti JC, Sakamoto-Hojo ET (2013) Lymphocytes of patients with Alzheimer's disease display different DNA damage repair kinetics and expression profiles of DNA repair and stress response genes. *International Journal of Molecular Sciences* 14: 12380-12400.
33. Carmona P, Molina M, Lopez-Tobar E, Toledano A (2015) Vibrational spectroscopic analysis of peripheral blood plasma of patients with Alzheimer's disease. *Anal Bioanal Chem* 407: 7747-7756.
34. Mordechai S, Shufan E, Porat Katz BS, Salman A (2016) Early diagnosis of Alzheimer's disease using infrared spectroscopy of isolated blood samples followed by multivariate analyses. *Analyst*.
35. Huleihel M, Shufan E, Zeiri L, Salman A (2016) Detection of vero cells infected with herpes simplex types 1 and 2 and varicella zoster viruses using raman spectroscopy and advanced Statistical methods. *PLoS One* 11: e0153599.
36. Salman A, Sebbag G, Argov S, Mordechai S, Sahu RK (2015) Early detection of colorectal cancer relapse by infrared spectroscopy in "normal" anastomosis tissue . *J Biomed Opt* 20: 75007.
37. Salman A, Shufan E, Lapidot I, Tsrer L, Moreh R, et al. (2015) Assignment of Colletotrichum coccodes isolates into vegetative compatibility groups using infrared spectroscopy: A step towards practical application . *Analyst* 140: 3098-3106.
38. Pomerantz A, Cohen Y, Shufan E, Ben-Naim Y (2014) Characterization of Phytophthora infestans resistance to mefenoxam using FTIR spectroscopy. *J Photochem Photobiol B* 141: 308-314.
39. Salman A, Shufan E, Zeiri L, Huleihel M (2014) Characterization and detection of vero cells infected with herpes simplex virus type 1 using Raman spectroscopy and advanced statistical methods. *Methods* 68: 364-370.
40. Fabian H, Jackson M, Murphy L, Watson PH, Fichtner I (1995) A comparative infrared spectroscopic study of human breast tumors and breast tumor cell xenografts . *Biospectroscopy* 1: 37-45.
41. Dovbeshko GI, Gridina NY, Kruglova EB, Pashchuk OP (2000) FTIR spectroscopy studies of nucleic acid damage. *Talanta* 53: 233-246.